Genetics of pathogenicity in Zymoseptoria tritici

Thierry MARCEL – University Paris-Saclay, INRAE UR BIOGER, France

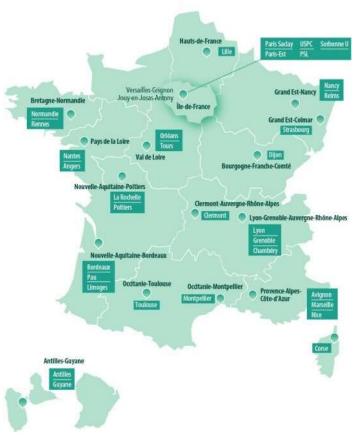




INRAE

National Research Institute for Agriculture, Food and Environment

18 Research Centres 14 Scientific Departments >200 Research Units >12,000 Employees



Our overarching goal is to address societal expectations for sustainable and environmentally friendly strategies in the fight against fungal diseases in major crop plants, and to manage risks associated with the dissemination and adaptive potential of phytopathogenic fungi.



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BIOGER Campus Agro Paris-Saclay



Reference research unit on fungal diseases of major European crops:

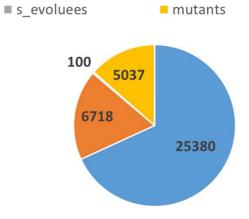


Inventory of 11 collections of fungal isolates:

Collections	Réfèrents	Genres concernés
AMAR	Clémentine Duplaix / Anne Sophie Walker	Botrytis Fusarium Microdochium Sclerotinia Ustilago Zymoseptoria Pyrenophora
BiogeR-syst	Valérie Laval / Noémie Vignolles	Fusarium Microdochium Parastagonospora
Botrytis	Muriel Viaud / Pascal Le Pêcheur	Botrytis
ECCP_Colleto	Richard O'Connell	Colletotrichum
EGIP	Anne Genissel / Marc-Henri Lebrun	Zymoseptoria
Epidémio_FS	Frédéric Suffert / Sandrine Gélisse	Zymoseptoria
Epidémio_HG	Henriette Goyeau	Puccinia
Epidémio_LG	Lilian Gout	Zymoseptoria Parastagonospora
Epidemio_ML	Marc Leconte	Puccinia
Epidemio_TM	Thierry Marcel	Zymoseptoria
EPLM	Mylène Balesdent / Laurent Coudard / Michel Meyer / Isabelle Fudal / Bénédicte Ollivier / Jessica Soyer / thésards	Leptosphaeria

Survey 2021 by Noémie Vignolles

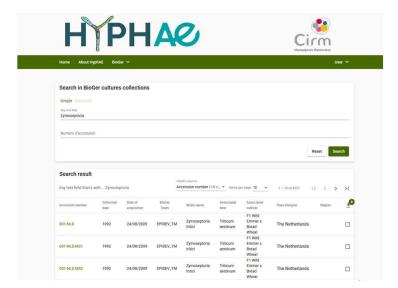
37.235 strains



s_issues_croisements

s_sauvages

Collaboration with CIRM Filamentous Fungi Marseille





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BIOINFOBIOGER: BIOGER Bioinformatics platform

https://bioinfo.bioger.inrae.fr/

Genomic resources for phytopathogenic fungi

BioinfoBioger is a platform dedicated to the management and analysis of genomic data for plant pathogenic fungi. It is part of the BIOGER unit, within INRAE's Plant Health and Environment division.

The platform is involved in multiple projects producing fungal genomes and populations omics data. The web portal provides access to different tools to explore these genomes and populations data through genome browsers or dedicated tools, and a data browser with raw and processed files and their related metadata. We also provide access to bioinformatics tools developed as part of our projects.

List of scientific <u>publications/productions</u> involving the members of the platform.

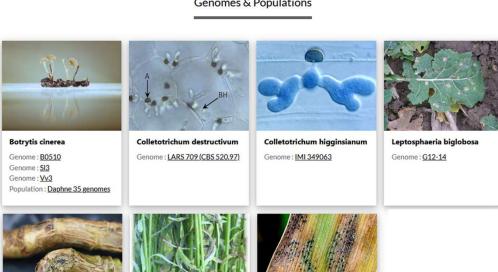






LAPALU Nicolas SIMON Adeline

Genomes & Populations



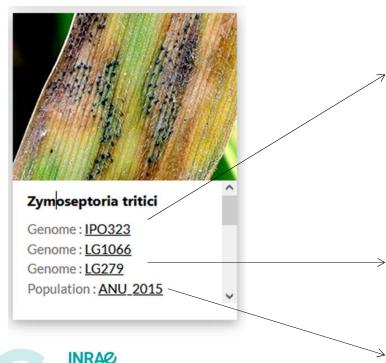


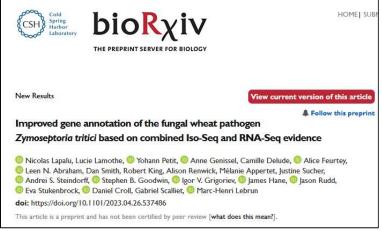
Leptosphaeria maculans

Genome: JN3

BIOINFOBIOGER: BIOGER Bioinformatics platform

https://bioinfo.bioger.inrae.fr/







LAPALU Nicolas SIMON Adeline

Two reference genomes of *Z. tritici* isolates specialized on durum wheat: INRA13-LG0279 & INRA13-LG1066

Whole genome Illumina sequences for 768 *Z. tritici* isolates (mostly French and European isolates), 321 of which have been pathotyped

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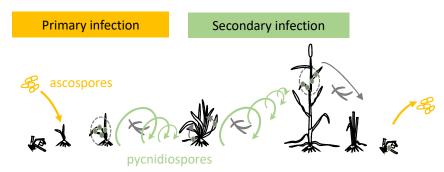
Biology of *Zymoseptoria tritici*

Cause the Septoria tritici blotch disease on wheat:

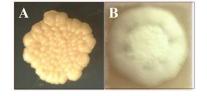


- Ascomycete (Dothideales)
- Dimorphic (yeast, mycelium)
- Heterothallic (Mat1, Mat2)
- Latent necrotroph

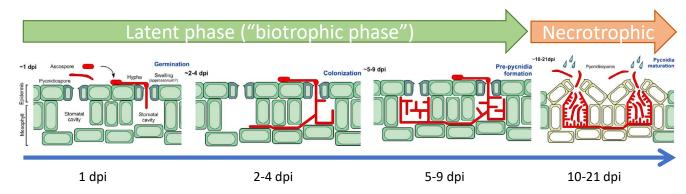




Development of a Septoria tritici blotch epidemic (Suffert et al., 2015)



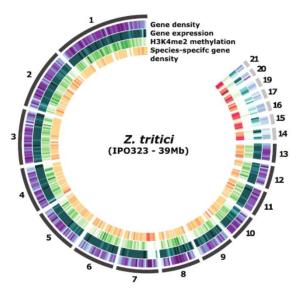
Dimorphism of *Z.tritici*:
A: yeast form
B: mycelium form
(Picture by Solweig LUCE)



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Plant infection stages of *Zymoseptoria tritici* (Steinberg, 2015)

Genetics of *Zymoseptoria tritici*



Feurtey et al. 2020

- Haploid (n)
- Genome size of 39.7 Mbp
- 13 core and 8 accessory chromosomes
- 13,414 re-annotated gene models

Controlled crosses between isolates

High level of

genetic diversity

"We found that wheat fields experiencing typical levels of infection are likely to carry

between 3.1 and 14.0 million pathogen genotypes per hectare and produce at least

2.1–9.9 trillion pycnidiospores per hectare."



Received: 14 March 2022 Revised: 28 May 2022 Accepted: 6 June 2022

ORIGINAL ARTICLE

Evolutionary Applications WILE

How large and diverse are field populations of fungal plant pathogens? The case of *Zymoseptoria tritici*

¹Plant Pathology Group, Institute of Integrative Biology, ETH Zurich, Zurich, Switzerland ²Université Paris-Saclay, INRAE, UR

Pathogen populations differ in the amount of genetic diversity they contain

McDonald et al. 2022



Orellana-Torrejon et al. 2022

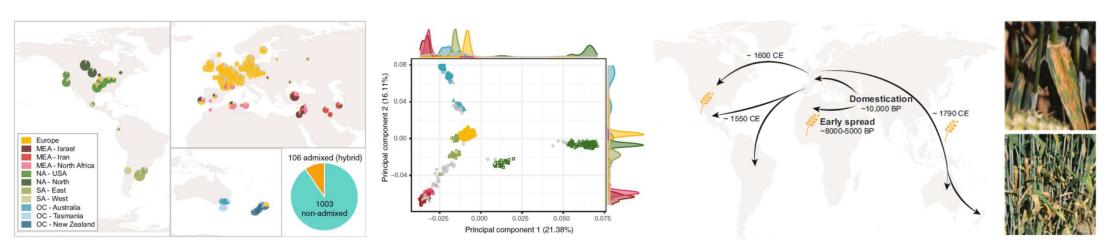
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Population genomics of *Z. tritici*

Worldwide genetic structure based on 1109 genomes:

Feurtey et al. (2023), Nature Communications 14





Eleven genetic clusters with differentiation mostly at the continental level:

 global genetic structure of the pathogen reveals multiple founder events associated with the introduction of wheat to new continents.

Global expansion of *Z. tritici* retracing the historical spread of its host:

- increased activity of transposable elements,
- significant standing variation for adaptation to new climates encountered during the global spread.

Genetics of pathogenicity in Z. tritici

Three reasons why it is crucial to identify pathogenicity factors in crop fungal pathogens:



1. Understanding Mechanisms of the Infectious Process

- How the pathogen infects, colonizes, and damages the host?
- Key molecular mechanisms such as effector functions, toxin production, and immune evasion.

2. Enhancing Durable Resistance in Crops [Effector Assisted Breeding]

- Use knowledge on fungal effectors to identify new resistance genes, and to help cloning.
- Develop wheat varieties with stronger and more durable resistance.
- Engineer disease resistance genes.

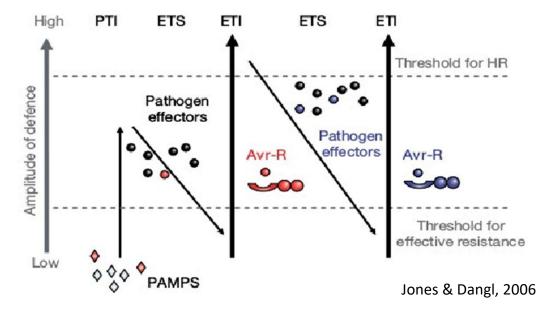
3. Monitoring and Predicting Pathogen Evolution

- Track virulent strains and understand how fungal populations adapt to host resistances.
- Forecast future outbreaks and emerging threats.
- Improve the strategic deployment of resistance genes.



Genetics of pathogenicity in Z. tritici

The zig-zag model in plant-pathogen interactions:



« Detection of an effector by an R protein triggers rapid activation of a very effective defense. »

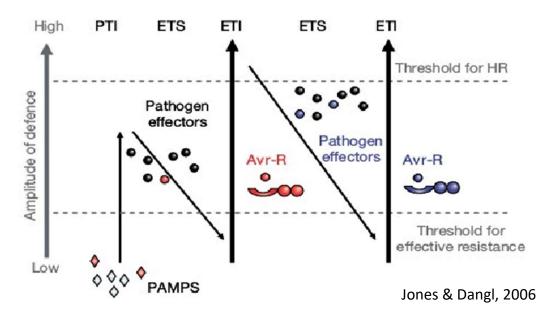
Pathogen effectors: molecule, typically a protein or small RNA, secreted by a fungal pathogen to manipulate the host plant's cellular processes and facilitate infection.

Avirulence genes: gene in a pathogen that encodes a product, typically an effector protein, that can be recognized by a corresponding resistance (R) gene in the host plant, triggering a defense response.



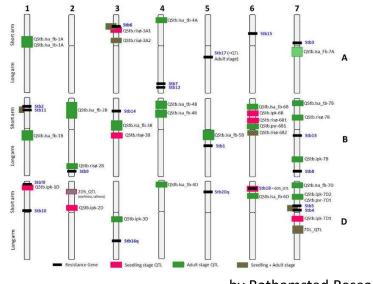
Genetics of pathogenicity in Z. tritici

The zig-zag model in plant-pathogen interactions:



« Detection of an effector by an R protein triggers rapid activation of a very effective defense. »

Genetic map of resistances to *Z. tritici* in bread wheat:



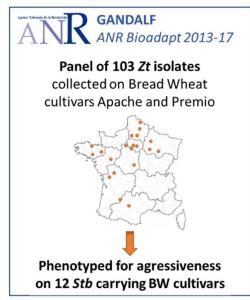
by Rothamsted Research

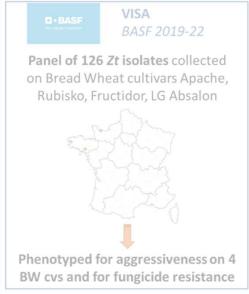
23 Stb resistance genes and 100s of QTLs

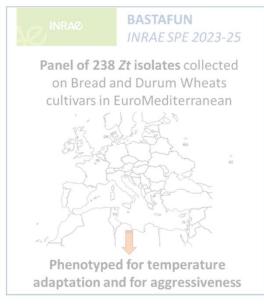
Are only *Stb* resistance genes involved in gene-for-gene interactions?

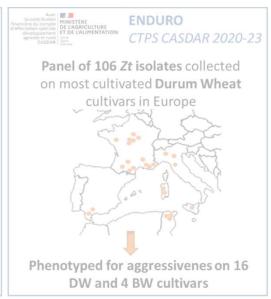


Dissection of host-pathogen interactions using GWAS







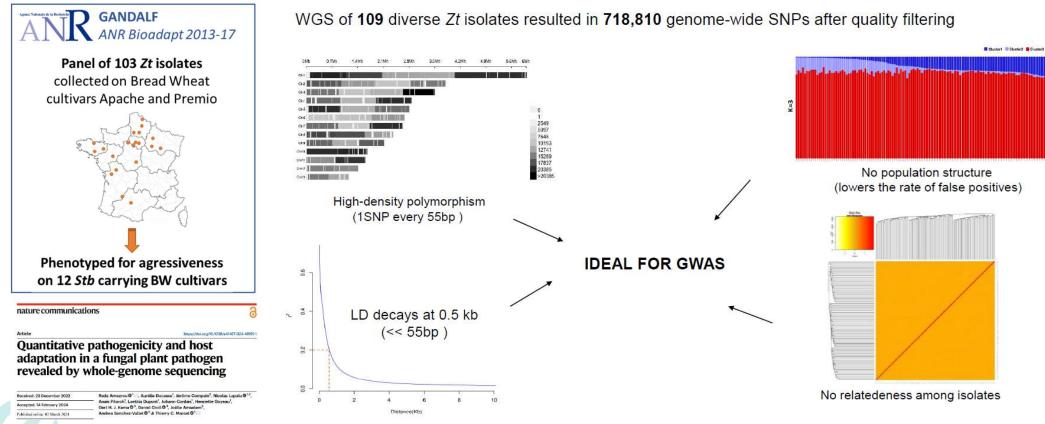




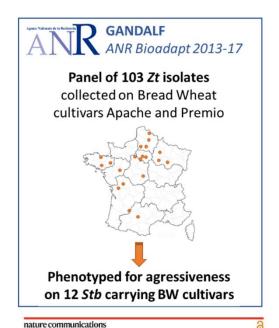
ceived: 23 December 2022
Reda Amezrou O** , Aurelle Ducasse*, Aerône Compain*, Nécolas Lapalu O**,
Ansis Pitarch, Lastitis Dapont , Johann Contias*, Herristette Goyesu*,
Gert H. J. Kens O**, Describ Coroll O**, Belle Amesian*,
Andrea Sancher-Vallet O** & Thierry C. Marcel O**

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Dissection of host-pathogen interactions using GWAS

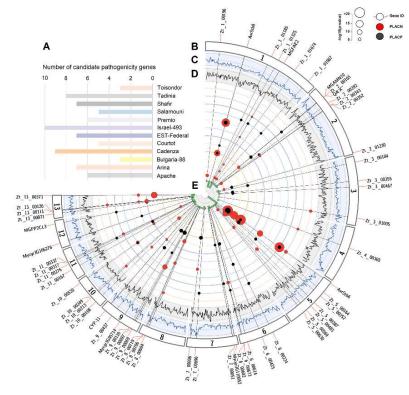


Dissection of host-pathogen interactions using GWAS



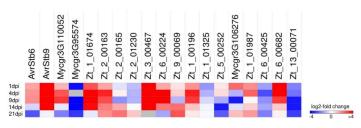






Amezrou et al. (2024), Nature Communications

Detection of 69 candidate genes, including nineteen genes highly expressed and/or differentially expressed in planta:



- > Pathogenicity in *Z. tritici* has a complex polygenic architecture and is predominantly quantitative.
- > Virulence-associated genes are mostly host specific.

Cloning of *AvrStb6*, responsible for avirulence on wheat cultivars carrying the resistance *Stb6*

AvrStb6 encodes a small secreted protein of 63 amino acids and rich in 11 cysteine residues

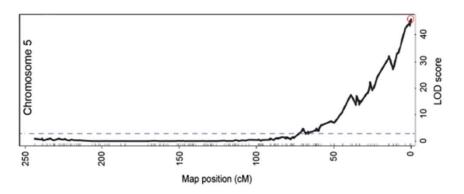


> The *Stb6-AvrStb6* interaction lead to a strong and asymptomatic disease resistance reaction

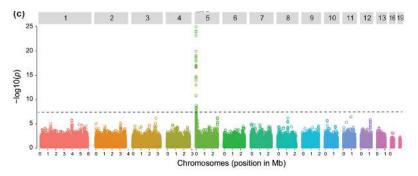
Zhong *et al.* (2017), New Phytologist



- Identified by QTL mapping

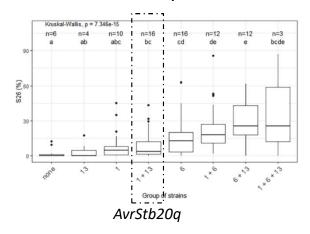


Identified by GWAS



Cloning of *AvrStb20q*, responsible for avirulence on wheat cultivars carrying the resistance *Stb20q*

AvrStb20g encodes a small secreted protein of 78 amino acids and rich in 11 cysteine residues

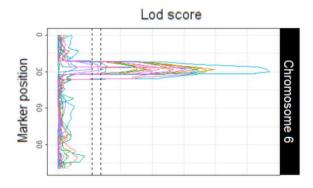


> The Stb20q-AvrStb20q interaction lead to a quantitative reduction in disease symptoms

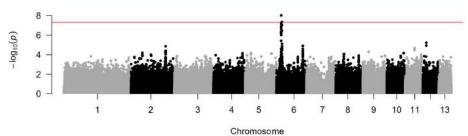
Langlands-Perry et al. (2023), Frontiers in Plant Science



Identified by QTL mapping

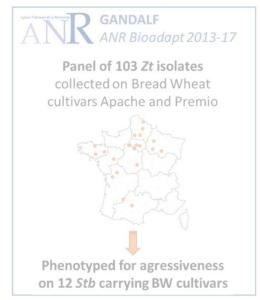


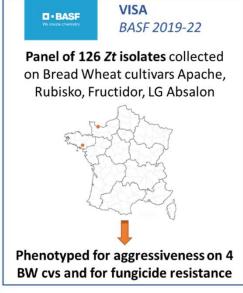
Identified by GWAS

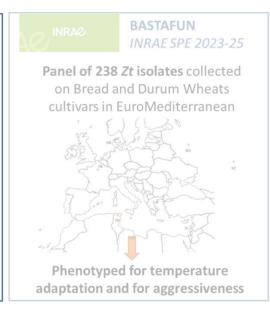


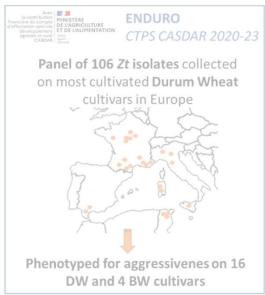
AUDPCS

Dissection of host-pathogen interactions using GWAS



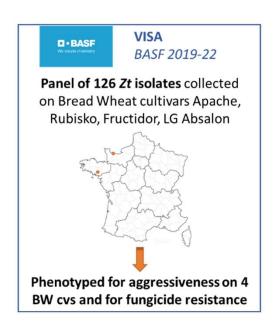




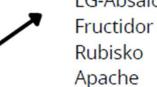




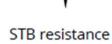
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Aggressiveness on the 4 wheat cultivars in controlled conditions



LG-Absalon

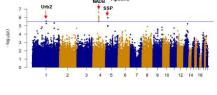


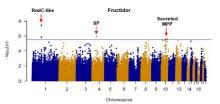


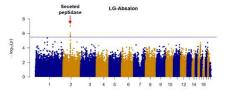
Percentage of leaf surface covered by:

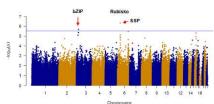
- necrosis
- pycnidia

Measured at 14, 20, 26 days post inoculation





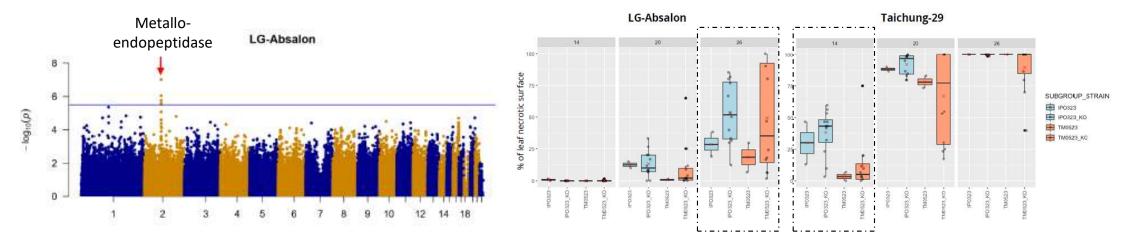




> Detection of 11 candidate genes



Cloning of a **metallo-endopeptidase** involved in *Z. tritici* pathogenicity

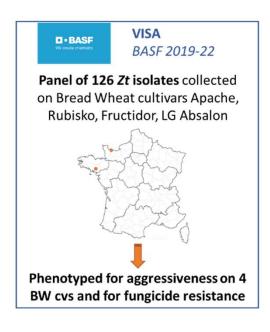


The peptidase-encoding gene was identified by GWAS only on the cultivar LG Absalon

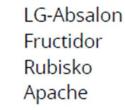


> The deletion of the peptidase-encoding gene tends to increase aggressiveness of *Z. tritici* independently of the cultivar





Aggressiveness on the 4 wheat cultivars in controlled conditions





STB resistance



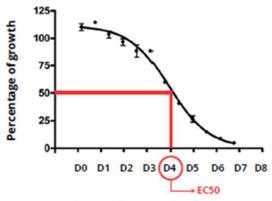
Percentage of leaf surface covered by:

- necrosis
- pycnidia

Measured at 14, 20, 26 days post inoculation

Fungicide resistance

4 DMIs (demethylation inhibitors) 2 SDHIs (succinate dehydrogenase inhibitors)

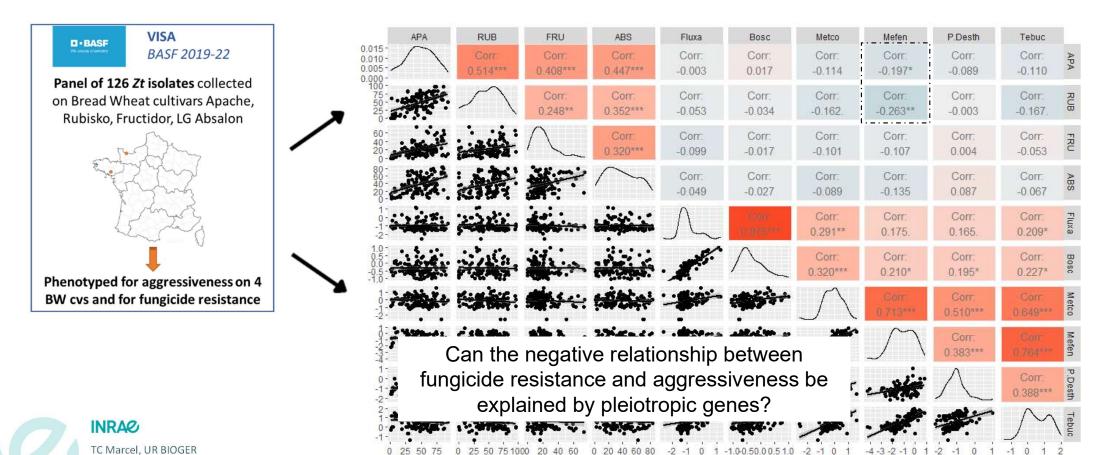


Fungicide concentration

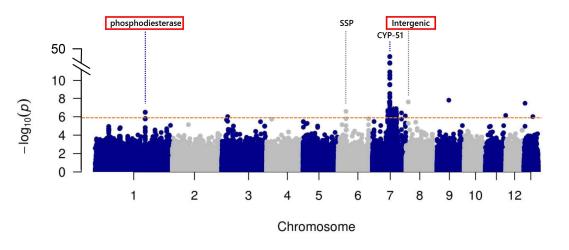
Adapted from Hanane Jary

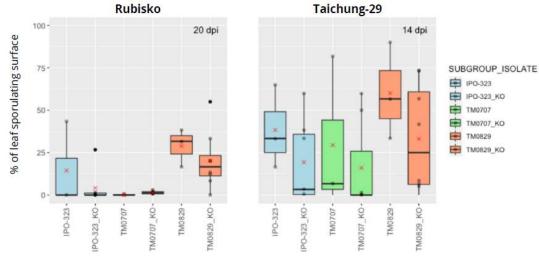
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Construction of a correlation matrix between aggressiveness and fungicide resistance traits:



The metaCCA algorithm (Cichonska *et al.*,2016) allows a powerful univariate genotype multivariate phenotype assocation testing, which is useful for correlated traits.





Phosphodiesterases (PDE) are regulators of cyclic nucleotide signaling (cAMP and cGMP). In fungi, PDE was shown to be involved in metabolism, stress responses, azole sensitivity and virulence etc.



> The PDE gene acts as a quantitative pathogenicity factor and is potentially involved in fungicide sensitivity in *Z. tritici*.



Take-home messages



In the wheat – Z. tritici pathosystem,

- Pathogenicity has a complex polygenic architecture and is predominantly quantitative
- Gene-for-gene interactions can underlie qualitative (AvrStb6) and quantitative (AvrStb20q) interactions
- Fungal populations exhibit standing genetic variation in pathogenicity genes (metalloendopeptidase) associated with aggressiveness, regardless of the host cultivar
- A phosphodiesterase gene may have antagonistic pleiotropic effects constraining adaptation of the fungus



Perspectives



- Validation of new avirulence and pathogenicity genes on-going...
- Construction of a pangenome graph for Z. tritici to improve the detection of candidate genes
- GWAS for pathogenicity of Z. trititi on durum wheat (PhD thesis)
- Allelic diversity of known Stb and AvrStb genes (PhD thesis)
- Development of high-throughput tools to follow the evolution of pathogenicity genes in fungal populations



Thank you...

Funders





VISA (2018-22)



DivR (2019-22) ATTILA (2023-26)

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Adaptive and epidemiological processes in wheat-fungal pathogen interactions



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